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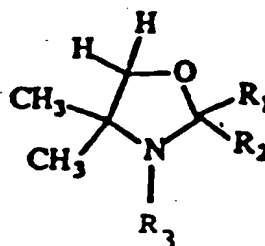
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(21) International Application Number: PCT/US96/09524 (22) International Filing Date: 7 June 1996 (07.06.96) (30) Priority Data: 08/473,960 7 June 1995 (07.06.95) US (71) Applicant: THE UNITED STATES OF AMERICA, represented by THE SECRETARY, DEPARTMENT OF HEALTH AND HUMAN SERVICES National Institute of Health [US/US]; Office of Technology Transfer, Suite 325, Rockville, MD 20852 (US). (72) Inventors: MITCHELL, James, B.; 10516 Sweepstakes Road, Damascus, MD 20872 (US). SAMUNI, Amran; 27 Even-Shmuel Street, Ramot, Jerusalem 97230 (IL). DEGRAFF, William, G.; 12020 Berry Street, Wheaton, MD 20902 (US). HAHN, Stephen; 5791 Trailwood Drive, Santa Rosa, CA 95404 (US). (74) Agents: GAGALA, Bruce, M. et al.; Leydig, Voit & Mayer, Ltd., Two Prudential Plaza, Suite 4900, 180 North Stetson, Chicago, IL 60601-6780 (US).		(81) Designated States: AL, AM, AT, AU, AZ, BB, BG, BR, BY, CA, CH, CN, CZ, DE, DK, EE, ES, FI, GB, GE, HU, IS, JP, KE, KG, KP, KR, KZ, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, TJ, TM, TR, TT, UA, UG, UZ, VN, ARIPO patent (KE, LS, MW, SD, SZ, UG), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG). Published <i>With international search report. Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.</i>	

(54) Title: NITROXIDES AS PROTECTORS AGAINST OXIDATIVE STRESS

(57) Abstract

The instant invention is directed to the use of a biologically compatible composition, containing an effective amount of a metal-independent nitroxide compound which is preferably represented by formula (I), wherein R₁ is -CH₃; R₂ is -C₂H₅, -C₃H₇, -C₄H₉, -C₅H₁₁, -C₆H₁₃, -CH₂-CH(CH₃)₂, -CHCH₃C₂H₅, or -(CH₂)₇-CH₃, or wherein R₁ and R₂ together



form spirocyclopentane, spirocyclohexane, spirocycloheptane, spirocyclooctane, 5-cholestane, or norbornane, R₃ is -O- or -OH, or a physiologically acceptable salt thereof, and a pharmaceutically acceptable carrier, as antioxidants capable of protecting cells, tissues, organs, and whole organisms against the deleterious effects of harmful free radical species generated during oxidative stress.

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NITROXIDES AS PROTECTORS AGAINST OXIDATIVE STRESS

BACKGROUND OF THE INVENTIONField of the Invention

The present invention relates to pharmaceutical compositions containing nitroxide compounds useful in ameliorating the deleterious effects of toxic oxygen-related species in living organisms, and methods of using the same.

Description of Related Art

The utilization of oxygen by mammals carries both a blessing and a potential curse. The blessing is that all mammals require oxygen for life. The potential curse is that during the metabolism of oxygen, a variety of toxic oxygen-related species such as hydroxyl radical, ($\cdot\text{OH}$); hydrogen peroxide, (H_2O_2); and superoxide, ($\text{O}_2\cdot$) are produced. Left unchecked, these free radical species could undoubtedly damage cells. However, cells have evolved elaborate detoxification and repair systems to rid themselves of these potentially toxic and undesirable metabolic by-products: superoxide dismutase (SOD) can convert superoxide to H_2O_2 , and catalase (CAT) can convert H_2O_2 to H_2O .

Yet another means to detoxify H_2O_2 (and organoperoxides) is via the enzyme glutathione peroxidase (GPX), which with glutathione (GSH), also converts H_2O_2 to H_2O . Glutathione transferase (GST), in addition to its ability to conjugate and inactivate drugs and xenobiotics, also possesses peroxidase activity and can detoxify H_2O_2 . These systems represent the major detoxification pathways for oxygen-derived free radicals species; however, there are doubtless

other systems that may provide protection including protein sulfhydryls and other thiol-related enzymes that could be involved in repair mechanisms.

Despite the efficiency of these enzymatic systems, there is a small "leakage" of toxic species beyond the biochemical defense network. Of particular importance is the ultimate fate of H_2O_2 , should it escape detoxification. H_2O_2 , itself an oxidant capable of damaging biologically important molecules, can also undergo reduction via ferrous complexes to produce $\cdot OH$. This reaction (often referred to as Fenton chemistry) produces the highly reactive $\cdot OH$, which in the order of 10^{-9} seconds, can: 1) abstract electrons from organic molecules; 2) break chemical bonds; 3) initiate lipid peroxidation; and 4) react with another $\cdot OH$ to produce H_2O_2 . It is not known whether chronic exposure to low level oxygen-derived free radical species is deleterious; however, it is postulated that the process of aging may be a manifestation of the organisms's inability to cope with sustained oxidative stress. Many modalities used in cancer treatment including x-rays and some chemotherapeutic drugs exert their cytotoxicity via production of oxygen-related free radicals, thereby imposing an added burden to normal detoxification systems. Additionally, free radicals and toxic oxygen-related species have been implicated in ischemia/reperfusion injury, and have long been thought to be important in neutrophil-mediated toxicity of foreign pathogens. Likewise, free radical damage has been implicated in carcinogenesis. The term "oxidative stress" has thus emerged to

encompass a broad variety of stresses, some of which have obvious implications for health care.

There has been considerable interest in devising additional approaches, apart from inherent intracellular detoxification systems, to protect cells, tissues, animals, and humans from the toxic effects of any agent or process that imposes oxidative stress. In the past few years, experimental studies have indicated that enzymes such as catalase and superoxide dismutase, and agents such as allopurinol and metal chelating compounds, afford protection against oxidative stress. None of these approaches is at present being applied to humans.

The application of the nitroxides of the instant invention is novel in this respect, and affords several unique advantages. Although the group of chemical compounds called stable nitroxide spin labels has had extensive biophysical use, they have never been used as antioxidants. They exhibit low reactivity with oxygen itself. Being low molecular weight, uncharged, and soluble in aqueous solution, they readily cross into the intracellular milieu. Enzymes such as catalase and superoxide dismutase do not. Therefore, the nitroxides should be superior to catalase and superoxide dismutase in that they can exert protection inside the cell. They are active within the biological pH range of about 5 to 8. Nitroxides are not proteins; therefore, the possibility of antigenic stimulation is remote. Previous low molecular weight superoxide dismutase mimics have all been metal dependent. The current agents do not contain metals, and problems with

dissociation constants and deleterious metal induced reactions are therefore avoided. These compounds are apparently non-toxic at effective concentrations, and their lipophilicity can be controlled by the addition of various organic substituents, facilitating targeting of the molecules to specific organs or organelles where toxic oxygen-derived species are generated, to regions which are particularly susceptible to oxidative damage, or to the brain, if this is so desired. Previous radiation protectors have been sulfhydryl-group dependent. The current agents do not have a sulfhydryl group. Finally, previous use of these types of compounds as magnetic resonance contrast agents does not relate to their instant application as antioxidants.

SUMMARY OF THE INVENTION

Accordingly, it is an object of the present invention to overcome the problems associated with the use of impermeable enzymatic detoxifying agents such as superoxide dismutase and catalase to protect living tissues from the deleterious effects of toxic products generated during oxygen metabolism. This is accomplished by providing a pharmaceutical composition containing the nitroxide compounds by any means, and methods for using the same as metal independent, low molecular weight antioxidant for use as:

- 1) Ionizing radiation protectants to protect skin, and to protect against mucositis, the effects of whole body radiation, and radiation-induced hair loss. Administration in these situations

may be accomplished either via topical application as an ointment, lotion, or cream, intravenously or orally by pill or lozenge.

2) Protectants against increased oxygen exposures so as to avoid, for example, pulmonary adult respiratory distress syndrome (ARDS).

3) Protectants against oxygen-induced lenticular degeneration and hyaline membrane disease in infants, and against oxidative stress-induced cataracts. The compounds may also be used to protect against oxidative stress in patients undergoing oxygen therapy or hyperbaric oxygen treatment. Administration under these circumstances may be accomplished via various routes including, for example, the use of eye drops, aerosol inhalation, or intravenous injection.

4) Reperfusion injury protectants effective in treating cardiovascular phenomena such as myocardial infarction and strokes, pancreatitis, or intestinal ulceration; to protect patients receiving organ transplants, and in organ preservation solutions.

5) Protectants for use in animal or plant cell culture media to prevent cytotoxicity due to excessive oxidation, for use in media designed for culturing aerobic microorganisms, for use in stabilizing labile chemical compounds which undergo spontaneous degradation by generating free radicals, for use in neutralizing free radicals which catalyze chain elongation during polymer formation, thereby terminating polymer elongation, and for use as a stabilizer for foods or food additives such as colors and flavors, especially in foods preserved via radiation treatment.

6) Biological antioxidants to protect humans and animals against agents such as the herbicide paraquat. In this circumstance, the pharmaceutical composition may be administered, for example, via inhalation as an aerosol to a subject exposed to paraquat. In addition, plants may be protected against such agents by, for example, spraying before or after exposure to such compounds.

7) Protectants against the cytotoxic effects of chemotherapeutic agents.

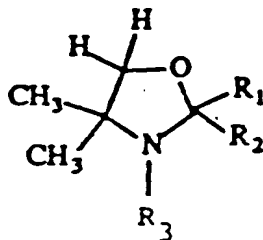
8) Protectants against mutagenic and carcinogenic agents. Administration in this situation or in 6, above, may be accomplished via oral ingestion, or parenterally.

9) Anti-inflammatory agents effective against arthritic conditions. For this purpose, the compositions may be administered parenterally, intra-articularly, or via oral ingestion.

10) Aging retardants. Administration for this purpose may be accomplished orally such as via a tablet supplement to the diet or parenterally.

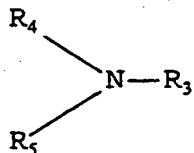
11) Oral or intravenous agents inducing weight reduction.

These and other objects are accomplished by providing a biologically compatible composition, comprising: an effective amount of a compound of the formula



wherein R_1 is $-CH_3$; R_2 is $-C_2H_5$, $-C_3H_7$, $-C_4H_9$, $-C_5H_{11}$, $-C_6H_{13}$, $-CH_2-CH(CH_3)_2$, $-CHCH_3C_2H_5$, or $-(CH_2)_7-CH_3$, or wherein R_1 and R_2 together form spirocyclopentane, spirocyclohexane, spirocycloheptane, spirocyclooctane, 5-cholestane, or norbornane; R_3 is $-O\cdot$ or $-OH$, or a physiologically acceptable salt thereof which has antioxidant activity; and a biologically acceptable carrier.

Compounds which may be useful in the present invention also include any compound having a $\text{>N-O}\cdot$ or >N-OH group, or a salt thereof. These compounds can be represented broadly by the formula:



wherein R_3 is as defined above, and R_4 and R_5 combine together with the nitrogen to form a heterocyclic group. The atoms in the heterocyclic group (other than the N atom shown in the formula) may be all C atoms or may be C atoms as well as one or more N, O and/or S atoms. The heterocyclic group preferably has 5 or 6 total atoms. The heterocyclic group may be preferably a pyrrole, imidazole, oxazole, thiazole, pyrazole, 3-pyrroline, pyrrolidine, pyridine, pyrimidine, or purine, or derivatives thereof, for example.

Further compounds which may be useful in the present invention also include those wherein R_4 and R_5 themselves comprise a substituted or unsubstituted cyclic or heterocyclic groups.

Still further compounds which may be useful in the present invention also include oxazolidine compounds capable of forming an oxazolidine-1-oxyl.

Yet further compounds which may be useful in the present invention also include metal-independent nitroxides.

The present invention is also directed to methods for treating the deleterious effects of harmful oxygen-derived metabolic products, as listed above.

Physiologically acceptable salts include acid addition salts formed with organic and inorganic acids, for example, hydrochlorides, hydrobromides, sulphates such as creatine sulphate salts, phosphates, citrates, fumarates and maleates. The compounds of the invention have been shown to exhibit little or no in vitro cytotoxicity at concentrations of from 1-5 mM for 5 hours.

The compounds of the present invention can be used for the treatment of the toxic effects of oxidative stress in a variety of materials, cells and mammals including humans, domestic and farm animals, and laboratory animals such as hamsters, mice, rats, monkeys, etc. It is contemplated that the invention compounds will be formulated into pharmaceutical compositions comprising an effective antioxidant amount of the compounds of formula (I) and pharmaceutically acceptable carriers. An effective antioxidant amount of the pharmaceutical composition will be administered to the subject or organism, human, animal, or plant, in a manner which prevents the manifestations of oxidative stress. The amount of the compound (I) and the specific pharmaceutically acceptable carrier

will vary depending upon the host and its condition, the mode of administration, and the type of oxidative stress condition being treated.

In a particular aspect, the pharmaceutical composition comprises a compound of formula (I) in effective unit dosage form. As used herein the term "effective unit dosage" or "effective unit dose" is denoted to mean a predetermined antioxidant amount sufficient to be effective against oxidative stress in vitro or in vivo. Pharmaceutically acceptable carriers are materials useful for the purpose of administering the medicament, which are preferably non-toxic, and may be solid, liquid or gaseous materials which are otherwise inert and medically acceptable, and are compatible with the active ingredients. The pharmaceutical compositions may contain other active ingredients such as antimicrobial agents and other agents such as preservatives.

These pharmaceutical compositions may take the form of a solution, emulsion, suspension, ointment, cream, aerosol, granule, powder, drops, spray, tablet, capsule, sachet, lozenge, ampoule, pessary, or suppository. They may be administered parenterally, intramuscularly or subcutaneously, intravenously, intra-articularly, transdermally, orally, buccally, as a suppository or pessary, topically, as an aerosol spray, or drops.

The compositions may contain the compound in an amount of from 0.1%-99% by weight of the total composition, preferably 1-90% by weight of the total composition, more preferably 10%-80% by weight of the total composition, and even more preferably, 25%-50% by

weight of the total composition. For intravenous injection, the dose may be about 0.1 to about 300 mg/kg/day. If applied topically as a liquid, ointment, or cream, the compound may be present in an amount of about 0.01 to about 100 mg/ml of the composition, more preferably in an amount of about 5 to about 75 mg/ml of the composition, and even more preferably in an amount of about 25 to about 50 mg/ml of the composition. For inhalation, about 0.1 to about 200 mg/kg body weight of the compound should be administered per day. For oral administration, the compound should be administered in an amount of about 0.1 to about 300 mg/kg/day.

The invention also provides a method for treating the effects of oxidative stress due to the production of harmful oxygen-derived species which comprises administering an effective antioxidant amount of the above-mentioned compound to a organism, material, mammal or human susceptible to oxidative stress. Such stress includes that due to oxidizing agents, increased oxygen exposure, oxygen-induced degeneration or disease, reperfusion injury, ionizing radiation, carcinogenic, chemotherapeutic, or mutagenic agents, aging, or arthritis.

Reperfusion injury may include myocardial infarction, strokes, pancreatitis, and intestinal ulceration, while oxidative stress due to increased oxygen exposure includes pulmonary adult respiratory distress syndrome. Other oxidative stresses amenable to treatment with the compounds of the instant invention include oxygen-induced lenticular degeneration, cataracts or hyaline membrane disease in infants, or oxidative stress occurring during oxygen therapy or

hyperbaric oxygen treatment.

Finally, in a further aspect of the invention, the compound of the instant invention can be used to prolong the storage life of human or animal cells, tissues, or organs by contacting these materials with a storage solution containing an effective amount of such compound, or to induce weight reduction in humans or animals.

The terms "biologically compatible" refer to a composition which does not cause any adverse effects to an organism to which it is applied. The composition is preferably free of toxic substances or other substances which would render it unsuitable for the intended use.

The term "parenteral" includes an administration by injection such as intravenous, intramuscular, or subcutaneous.

BRIEF DESCRIPTION OF THE DRAWINGS

FIGURE 1A-1D shows EPR spectra of CHD and TEMPO demonstrating the partitioning of each nitroxide (1 mM) in both the intra- and extra-cellular space of V79 cells. The EPR signal intensity of the total concentration of CHD (intra- and extra-cellular) in 6.4×10^7 V79 cells/ml traces (A and C) and in the presence of 110 mM trioxalato-chromiate (CrOx) (traces B and D). The gains for individual spectra are as cited in the individual figures.

FIGURE 2 shows a survival curve for Chinese hamster V79 cells exposed to HX/XO (hypoxanthine/xanthine oxidase) in the presence of various additives, including the nitroxides CHD and TEMPOL, which fully protected the cells. Chinese hamster V79 cells in full

medium at 37°C were exposed to 0.05 U/ml XO + 0.5 mM HX for various time periods in the presence of various additives: (), control, no additives; (■), 100 U/ml catalase; (O), 100 U/ml SOD; (□), 500 μ M DF (desferrioxamine), preincubated for 2h with the cells prior to addition of XO; (Δ), 5 mM CHD; (\blacktriangle), 5 mM TEMPOL.

FIGURE 3 shows a survival curve for cells exposed to H_2O_2 in the presence of various additives, including the nitroxides CHD and TEMPOL, which fully protected the cells. The effect of various agents on cell survival was measured by clonogenic assay of Chinese hamster V79 cells exposed in full medium at 37°C to various concentrations of H_2O_2 for 1h; (●), control, no additives; (■), 100 U/ml catalase; (O), 100 μ g/ml SOD; (□), 500 μ M DF, preincubated 2h with the cells prior to H_2O_2 addition; (Δ), 5 mM CHD; (\blacktriangle), 5 mM TEMPOL.

FIGURE 4 shows the effect of nitroxide on the accumulation and decay of H_2O_2 in a tissue culture of cells exposed to HX/HO. Chinese hamster V79 cells were plated in full medium and incubated at 37°C with 5 mM HX + 0.04 U/ml XO, sampled at various time points, and assayed for H_2O_2 using a hydrogen peroxide selective electrode.

FIGURE 5 shows the survival of Chinese hamster V79 cells exposed to 600 μ M H_2O_2 \pm DF or CHD in full medium at 37°C for 1h under hypoxic conditions.

FIGURE 6 shows the reaction between CHD and DNA-Fe(II): CHD in 50 mM MOPS buffer pH 7.0 was anoxically mixed at 22°C with DNA-Fe(II). All solutions always contained 0.1 mg/ml Salmon DNA. The appearance of DNA-Fe(III) was spectrophotometrically monitored at

353 nm, whereas the spin-loss of CHD was monitored by following its EPR signal. To study the time-dependence of ΔOD_{353nm} (O), 1 mM CHD was mixed with 0.1 mM Fe(II). To follow the spin-loss of CHD (\square), 1 mM Fe(II) was mixed with 0.1 mM CHD.

Inset: Time-dependence of $\ln\{\text{EPR signal}\}$ (\square); and of $\ln\{OD_0 - OD_t\}$ (O).

FIGURE 7 shows the clonogenic survival of Chinese hamster V79 cells treated with varying concentrations of TEMPOL 10 min prior to radiation.

FIGURE 8 shows the protection afforded whole animals treated with TEMPOL prior to whole body irradiation. Six week old female C3H mice were given 275 mg/kg TEMPOL intraperitoneally 10 mins prior to irradiation with 3 Gy to 13 Gy. Controls were given saline.

FIGURE 9 shows the average weights of control and TEMPOL treated mice.

FIGURE 10 shows the mean weight gain over 28 weeks post-irradiation for four groups of animals. Control = no treatment (closed triangles); animals receiving no radiation, but given 10 mg/ml TEMPOL in their drinking water for two weeks (open triangles); animals receiving 24 Gy to the kidneys (closed circles); animals receiving 24 Gy to the kidneys plus 10 mg/ml TEMPOL in their drinking water two weeks post-irradiation (open triangles).

FIGURE 11A shows a section from a kidney of a control animal not treated with radiation. Sections from kidneys of animals

receiving TEMPOL alone were identical to those of controls (data not shown).

FIGURE 11B shows a section of a kidney from a mouse treated with a single radiation dose of 24 Gy. Significant evidence of sclerotic glomeruli and tubular cell loss can be seen.

FIGURE 11C shows a section of a kidney from a mouse that received a single radiation dose of 24 Gy and orally administered TEMPOL for two weeks post-irradiation. Protection against glomerular sclerosis and tubular cell loss can be readily observed.

DESCRIPTION OF THE PREFERRED EMBODIMENTS

EXAMPLE 1

Synthesis and SOD-like Activity of Oxazolidine Derivatives

In Vitro

Desferrioxamine (DF) was a gift from Ciba Geigy; hypoxanthine (HX) was purchased from Calbiochem - Boehringer Co.; 2,2,6,6-tetramethylpiperidine-1-oxyl (TEMPO), 4-hydroxy-2,2,6,6-tetramethylpiperidine-1-oxyl (TEMPOL), 4-hydroxypyrazolo[3,4,-d] - pyrimidine (allopurinol), p-toluene sulfonic acid, 2-amino-2-methyl-1-propanol, 2-butanone, and cyclohexanone were purchased from Aldrich Chemical Co.; trioxalato chromiate(III) (CrOx) was purchased from Pfaltz and Bauer, Inc., and recrystallized; xanthine oxidase (EC 1.2.3.2. xanthine: oxygen oxidoreductase) grade III from buttermilk, superoxide dismutase (SOD), and grade V ferricytochrome c were obtained from Sigma. H₂O₂ was bought from

Fisher Scientific Co. XO was further purified on a G25 sephadex column. All other chemicals were prepared and used without further purification. Distilled-deionized water was used throughout all experiments.

CHD, 2-spirocyclohexane doxyl (2-spirocyclohexane-5,5-dimethyl-3-oxazolidinoxyl) and OXANO, 2-ethyl-2,5,5-trimethyl-3-oxazolidine-1-oxyl as well as other nitroxides were synthesized as described by Keana et al (J. Am. Chem. Soc., 89, 3055-3056, 1967). For the general synthesis of the cyclic amines, the appropriate starting ketone was reacted with 2-amino-2-methyl-1-propanol in benzene in the presence of catalytic amounts of p-toluene sulfonic acid. As the cyclic structure formed, water was eliminated. The volume of water collected in a Dean Stark apparatus was monitored and used to gauge the reaction progress. The amines thus produced were purified through fractional distillation under reduced pressure, characterized by 220 MHz ¹H NMR, IR, UV, either EI or CI mass spectroscopy, and subsequently oxidized to the corresponding nitroxides using m-chloroperbenzoic acid. The nitroxides were purified by silica flash chromatography (Still et al, J. Org. Chem., 43, 2923-2925, 1978). Water/octanol ratios were determined by placing a quantity of nitroxide in water+octanol within a separatory funnel. The mixture was shaken thoroughly and allowed to separate for 15 min, whereupon aliquots were taken from both fractions and the ratio of nitroxide distribution was determined using electron paramagnetic resonance (EPR) spectroscopy, by comparing the intensities of signal obtained under N₂.

To check whether oxazolidinoxyl derivatives other than OXANO manifest SOD-like activity, several nitroxides having different ring substituents were synthesized. Table 1A shows representative synthesized nitroxides with accompanying physical characteristics.

Table 1A. Five-Membered Oxazolidine-1-oxyl (Doxyl)

	Nitroxide notation	Ring substituents R ₁ R ₂		Yield (%)	Partition [#] coefficient
I	OXANO	CH ₃	C ₂ H ₅	42	10
II		CH ₃	C ₅ H ₁₁	52	145
III		CH ₃	C ₄ H ₉	49	58
IV	CHD	spirocyclohexyl		77	80
V		CH ₃	C ₆ H ₅	22	720

[#]Octanol:water

Exposure of these 5-membered cyclic nitroxides to O₂ flux formed by HX/XO resulted in a decrease in their EPR signal, as previously found for OXANO (Samuni et al, Free Rad. Biol. Med., 6, 141-148, 1989). For EPR experiments, samples (0.05-0.1 ml) either of solutions of chemicals or cell suspensions were drawn by a syringe into a gas-permeable teflon capillary of 0.8 mm inner diameter, 0.05 mm wall thickness (Zeus Industrial Products, Inc. Raritan, NJ). Each capillary was folded twice, inserted into a

narrow quartz tube which was open at both ends (2.5 mm ID), and then placed horizontally into the EPR cavity. During the experiments, gases of desired compositions were blown around the sample without having to disturb the alignment of the tube within the EPR cavity. EPR spectra were recorded in a Varian E4 (or E9) X-band spectrometer, with field set at 3357G, modulation frequency of 100 KHz, modulation amplitude of 1G and non-saturating microwave power. The EPR spectrometer was interfaced to an IBM-PC through an analog-to-digital converter and a data translation hardware (DT2801) and the spectra were digitized using commercial acquisition software, enabling subtraction of background signals. To study the kinetics of the spin-loss, the spectra were deliberately overmodulated, and the magnetic field was kept constant while the intensity of the EPR signal was followed.

After terminating the HX/XO reaction by allopurinol, the nitroxide spin-loss was reversed by adding 0.5 mM ferricyanide, indicating that O_2 reduces the nitroxide to its respective hydroxylamine (Samuni et al, Free Rad. Biol. Med., 6, 141-148, 1989). On the other hand, no effect of O_2 on the EPR signal of 6-membered ring nitroxides such as TEMPO and TEMPOL was detectable (see Table 1B).

Table 1B. Kinetic Data: SOD-like Activity of 5- and 6-Membered Cyclic Nitroxides

Chemical Structure

Nitroxide notation	TEMPOL	TEMPO	OXANO	CHD
Steady state EPR signal (%) ^a	100	100	50	30
$kR'RNO+O_2$ (M ⁻¹ s ⁻¹) ^b	3.4×10^5	1.3×10^6	1.1×10^3	3.5×10^3

^aSteady-state EPR signal of nitroxides (% from total R'RNO+R'RNOH) after exposure to 5 mM HX + 0.03 U/ml XO in air-saturated PBS pH 7.2.

^bRate constants were determined at low ionic strength (10 mM HEPES), pH 7.0, and 22°C.

The failure of superoxide to affect TEMPO and TEMPOL apparently suggested that 6-membered cyclic nitroxides lack SOD-like activity. As a further check, the reaction of representatives of both 5- and 6-membered cyclic nitroxides with O₂ was studied. The SOD-inhibitable ferricytochrome c reduction assay (Fridovich, Handbook of Methods for Oxygen Radical Research, 213-215, 1985) was used to determine rate constants of reaction with O₂. Superoxide radicals were generated at 25°C in aerated phosphate buffer (50 mM) containing 50 μM DTPA, 5 mM HX, and 10-50 μM ferricytochrome c (with or without 65 U/ml catalase). The reaction was started by adding 0.01 U/ml XO and the rate of ferricytochrome c reduction, in the absence (V) and in the presence (v) of various nitroxides, was

spectrophotometrically followed at 550 nm. Both reference and sample cuvettes contained all the reagents, with the reference cuvette containing 100 units/ml SOD, thereby eliminating spurious reactions from interfering with the determination of rate constants. Data were analyzed by plotting V/v as a function of [nitroxide] and k_1 was calculated, knowing $k_{\text{CytC}+\text{superoxide}}$, according to:

$$(V/v) - 1 = k_1 \times [\text{nitroxide}] / k_{\text{CytC}+\text{superoxide}} \times [\text{Cyt-c}^{\text{III}}].$$

Via this assay, all the nitroxides listed below have been shown to function as superoxide dismutase mimics.

Table 2. Physical data of 2-substituted-5,5-dimethyl-3-oxazolidines.

Generalized Structure, R groups shown in table.

Oxazolidine (A-I)	R1	R2	Yield%	b.p. C (760mmHg)
A	CH3	C2H5	42	162-165
C	CH3	C3H7	36	169-170
D	CH3	C4H9	71	185-188
E	CH3	C5H11	52	204-205
F	CH3	C6H13	54	225-230
G	CH3	CH2CH(CH3)2	49	168-170
H	CH3	CHCH3C2H5	25	184-185
I	CH3	(CH2)7CH3	57	165-166
Oxazolidine (J-R)	Alicyclic substituent		Yield%	b.p. C (760mmHg)
J	spirocyclopentane		61	133-135 (75)
K	spirocyclohexane		77	208-211 (760)
L	spirocycloheptane		50	145-147 (35)
M	spirocyclooctane		40	234-235 (760)
N	5-cholestane		--	--
O	norbornane		53	178-180 (80)

The rate constants of the synthetic nitroxides' reaction with O_2 at low ionic strength (10mM HEPES) and pH 7.0 ranged from 1.1×10^3 to $1.3 \times 10^6 M^{-1}s^{-1}$, as compared with $2.3 \times 10^9 M^{-1}s^{-1}$ for k_{cat} of native SOD.

None of the nitroxides shown (in the last two tables) exhibited cytotoxicity determined by clonogenic assay in V79 cells exposed for 1h at 5 mM. For subsequent studies, the most lipophilic nitroxide, CHD, and the most hydrophilic one, TEMPOL, were chosen.

EXAMPLE 2

Nitroxide Intracellular Localization

Figure 1A and C illustrate the EPR signal from 1 mM CHD and TEMPOL, respectively, suspended with 6.4×10^7 V79 cells/ml. This EPR signal represents the total concentration of intra- and extracellular CHD. Trioxalato-chromiate is a paramagnetic broadening agent which remains excluded from the intracellular volume space and causes the EPR signal from extracellular species to become non-detectable (Lai, Biophys. J., 52, 625-628, 1987). When cells were added to CHD or TEMPOL in the presence of 110 mM trioxalato-chromiate, a much smaller yet observable intracellular signal was detected as shown in Figure 1B and D. The observable line broadening and loss of the hyperfine structure of the intracellular signal indicate that CHD, though not the TEMPOL, has decreased freedom of motion (anisotropy) within the intracellular environment and is located primarily in a membranous compartment as can be anticipated based on the difference between their lipophilicities.

EXAMPLE 3Protection of Cells Against Oxidative Damage

Chinese hamster V79 cells were grown in F12 medium supplemented with 10% fetal calf serum, penicillin, and streptomycin. Survival was assessed in all studies by the clonogenic assay. The plating efficiency range between 80-90%. Stock cultures of exponentially growing cells were trypsinized, rinsed, and plated (5×10^5 cells/dish) into a number of 100 mm petri dishes and incubated 16h at 37°C prior to experimental protocols. Cells were exposed for 1h at 37°C to either 0.5 mM hypoxanthine (HX) + 0.05 U/ml of xanthine oxidase (XO) for varying lengths of time, or to H_2O_2 at different concentrations. To assess possible modulation in cytotoxicity, catalase, 100 U/ml; SOD, 100 μ g/ml; DF, 500 μ M; and 5 mM of each of the nitroxides from Table 1 were added to parallel cultures. CHD was prepared in a stock solution in ethanol and diluted into medium such that the final concentration was 5 mM. This resulted in a final concentration of 1% ethanol in the medium which was not cytotoxic and did not influence the cellular response to HX/XO or H_2O_2 . TEMPOL is water soluble and was prepared directly in tissue culture medium. Neither catalase, SOD, DF, CHD, nor TEMPOL were cytotoxic alone in the concentrations used. DF was added either 2h prior to or during treatment while the other agents were present only during HX/XO or H_2O_2 treatment. Following treatment, cells were rinsed, trypsinized, counted, and plated for macroscopic colony formation. For each dose determination cells were plated in triplicate and the experiments

were repeated a minimum of two times. Plates were incubated 7 days, after which colonies were fixed with methanol/acetic acid, stained with crystal violet, and counted. Colonies containing >50 cells were scored. Error bars represent S.D. of the mean and are shown when larger than the symbol.

Some studies required exposure to H_2O_2 under hypoxic conditions. For these studies, cells dispersed in 1.8 ml of medium were plated into specially designed glass flasks (Russo et al, Radiat. Res., 103, 232-239, 1985). The flasks were sealed with soft rubber stoppers, and 19-gauge needles were pushed through to act as entrance and exit ports for a humidified gas mixture of 95% nitrogen/5% CO_2 (Matheson Gas Products). Each flask was also equipped with a ground glass side arm vessel which when rotated and inverted could deliver 0.2 ml of medium containing H_2O_2 at a concentration which when added to the cell monolayer resulted in final concentration of H_2O_2 of 600 μM . Stoppered flasks were connected in series and mounted on a reciprocating platform and gassed at 37°C for 45 min. This gassing procedure results in an equilibrium between the gas and liquid phase (in both the medium over the cell monolayer and in the solution in the sidearm) and yielded oxygen concentrations in the effluent gas phase of <10 ppm as measured by a Thermox probe (Russo et al, Radiat. Res., 103 232-239, 1985). After 45 min of gassing, the hypoxic H_2O_2 solution was added to the cell monolayer culture. The cells were exposed to H_2O_2 for 1h under hypoxic conditions. N_2 gas flow was maintained during the H_2O_2 exposure. In parallel flasks, DF and CHD were added as

described above, and were present during the entire gassing procedure. Following treatment, cell survival was assessed as described above.

Hydrogen peroxide was assayed using a YSI Model 27 Industrial Analyzer (Yellow Springs Instruments) equipped with a selective electrode for H_2O_2 . For analysis of cellular preparations, the cells, except during the brief time required for removal of aliquots for analysis, were kept in T25 culture flasks maintained at 37°C in complete medium (pH 7.2). Aliquots of 25 μ l were sampled from the reaction or cell preparation system at varying time points and injected into the analyzer. $[H_2O_2]$ was determined after calibrating the instrument with known concentrations of H_2O_2 . The concentrations of standard H_2O_2 solutions were calibrated using iodometric assay (Hochanadel, J. Phys. Chem., 56, 587-594, 1952).

To expose the cells to oxidative stress they were incubated with HX/XO. Figure 2 shows a survival curve for cells exposed to HX/XO. Cell survival was not altered when SOD was present during the HX/XO exposure. In contrast, 5 mM CHD or TEMPOL fully protected the cells. The other nitroxides, presented in Table I, afforded similar protection (data not shown). Figure 2 also shows that either catalase or DF provides complete protection from HX/XO-derived damage. Complete protection by DF required a 2h preincubation with DF before cells were exposed to HX/XO, whereas DF addition simultaneously with HX/XO offered only partial protection (data not shown).

One interpretation of the data shown in Figure 2 is that H_2O_2

is the principal cytotoxic species produced by the HX/XO system (Link & Riley, Biochem. J., 249, 391-399, 1988). This assumption is based on the fact that extracellular catalase provided complete protection from HX/XO (Figure 2). To test if cell protection by the SOD-mimic resulted from detoxifying H_2O_2 , cells were exposed to H_2O_2 as shown in Figure 3. The results of these experiments were identical to those shown in Figure 2, in that SOD did not protect, but catalase, DF, TEMPOL, and CHD provided complete protection against H_2O_2 cytotoxicity. At this point it was questioned if CHD might have other features apart from acting as a SOD mimic, namely, whether CHD affects H_2O_2 concentration. Figure 4 shows the concentration of H_2O_2 in tissue culture exposed to HX/XO. With time there was a build-up followed by a slow decline in $[H_2O_2]$. The presence of CHD did not significantly alter the pattern of H_2O_2 generation by HX/XO. Thus, the cellular protection afforded by CHD to HX/XO and H_2O_2 could not be attributed to a direct reaction of CHD with H_2O_2 .

Even with direct exposure of cells to H_2O_2 , there is the possibility that superoxide could be produced intracellularly as a result of the H_2O_2 treatment. If superoxide were produced intracellularly, CHD protection of cells from HX/XO and H_2O_2 might be expected, given the findings that CHD can penetrate intracellular spaces as shown in Figure 1. To test if the cytoprotection provided by CHD was solely a result of its reaction with superoxide, CHD effectiveness was examined when H_2O_2 was applied to cells incubated in a hypoxic environment, conditions in

which the chance for superoxide formation would be significantly limited. As is seen in Figure 5, CHD protects against H_2O_2 cytotoxicity even under hypoxic conditions.

Figure 5 also shows that DF provides complete protection to H_2O_2 cytotoxicity under hypoxic conditions. The pattern of DF protection shown in Figures 2, 3, and 5 suggested that the cytotoxicity of HX/XO and H_2O_2 may directly involve intracellular reduction of H_2O_2 by ferrous ion to produce the highly toxic $\cdot OH$. It was also questioned whether the aerobic and hypoxic protection by CHD to H_2O_2 exposure was a result of CHD directly accepting electrons from ferrous ions, thereby preventing generation of $\cdot OH$. Because cellular iron is chelated, the possible reaction of nitroxide with chelated iron(II) was examined by repeating the experiment in the presence of DNA. To study the possibility of nitroxide-induced oxidation of transition metals, CHD was hypoxically mixed with iron(II) in the presence of 0.1 mg/ml Salmon DNA. Consequently, DNA-Fe(III) was formed and the nitroxide EPR signal disappeared. The reaction kinetics were investigated by maintaining either CHD or Fe(II) in excess while the absorbance due to DNA-Fe(III) and the nitroxide spin-loss were monitored respectively (Figure 6). Both the decay of the EPR signal and the appearance of the OD_{353nm} obeyed pseudo 1st order kinetics from which the 2nd order reaction rate constant was calculated as 44 $M^{-1}s^{-1}$ or 33 $M^{-1}s^{-1}$ using the data from EPR or optical absorption, respectively. When TEMPOL was hypoxically mixed with DNA-Fe(II), a similar reaction took place having a 2nd order reaction rate constant of 40

M¹s⁻¹. The spin-loss was completely reversed by adding 2 mM ferricyanide, thus indicating that DNA-Fe(II) reduced the respective nitroxide to its hydroxylamine.

EXAMPLE 4

In Vitro and In Vivo Protection

Against Ionizing Radiation by Nitroxide

Chinese hamster V79 cells were treated with varying concentrations of TEMPOL 10 min prior to irradiation. The clonogenic survival compared to control cells is shown in Figure 7. The extent of protection for 100 mM TEMPOL was approximately 2.5 fold.

The protection afforded whole animals by TEMPOL was evaluated in six week, old female C3H mice given 275 mg/kg TEMPOL intraperitoneally 10 min prior to whole body radiation doses ranging from 3 Gy to 13 Gy. Controls were given saline. Survival was recorded 30 days after exposure to radiation. The LD₅₀₍₃₀₎ refers to that dose of radiation at which 50% of the mice survived 30 days after exposure. As can be seen in Figure 8, mice treated with TEMPOL had an approximately 25% higher LD₅₀₍₃₀₎, demonstrating protection from whole body radiation and no toxicity.

The above results demonstrate that TEMPOL provides radio-protection at both the in vitro and in vivo level.

EXAMPLE 5

Nitroxide-Induced Weight Loss In Animals

Six week old female C3H mice (6 animals in each group) were allowed to drink an unlimited supply of water alone (control) or 4-hydroxy tempo (TEMPOL) dissolved in water at a concentration of 10 mg per ml. Chronic oral administration (>3 weeks) results in no apparent toxicity to the animals, but a reduction of weight compared to controls by 12.5% as shown in Figure 9. Thus, nitroxide given over extended periods appears to cause weight loss in animals.

EXAMPLE 6

Post-Ionizing Radiation Protection By Nitroxides

Against Cell, Tissue, And Organ Damage

Therapeutic radiation for cancer treatment is often compromised because normal cells, tissues, and organs are near, or surround, the tumor being treated. This poses a problem in that normal cells, tissues, and organs can only withstand a limited dose of ionizing radiation before permanent damage occurs. Unfortunately, this limited normal tissue dose is often inadequate to control or kill the tumor. Thus, any chemical intervention that facilitates selective radio-protection of normal tissue as compared to tumor tissue without damage to nearby cells would be beneficial in that a greater radiation dose could be delivered to the tumor, thereby enhancing the likelihood of tumor control.

It has been shown in rodents that nitroxides can provide

radio-protection to normal tissues while they do not protect tumors. Nitroxides can therefore significantly protect normal tissues, permitting delivery of higher ionizing radiation doses to tumors. This protective effect can be achieved by administering nitroxides immediately prior to, during, or after radiation exposure.

Radiation damage to normal tissues and organs is most probably due in large part to damage to cells comprising the tissue or organ and/or damage to blood vessels supplying the tissue or organ (Fajardo et al. (1988) Pathol. Ann. 23:297-330; Hopewell et al. (1986) Br. J. Cancer 53 (Suppl. VII):181-191). However, it has been known for many years that radiation causes inflammation in normal tissues and organs (Rubin et al. (1968) In Clinical Radiation Pathology, W.B. Saunders Company, Philadelphia, pp.38-61), most likely mediated by infiltration of host leukocytes. The clinical sequelae of this transient inflammation has not been exhaustively studied; however, in several clinical situations of chronic radiation injury, anti-inflammatory steroid agents have been shown to be of benefit. Leukocytes infiltrating irradiated tissue may become "activated" and undergo oxidative bursts that can produce free radicals, including oxygen-related free radicals such as superoxide, hydrogen peroxide, and hydroxyl radicals (Malech et al. (1987) The New England Journal of Medicine 317(11):687-694). Production of these species may well underlie the observed inflammation seen in normal tissues and organs following radiation treatment; however, they may also contribute to the damage process

of tissues and organs as well. Damage to organs as a result of "activated" leukocytes has been reported by Tate et al. ((1984) In Free Radicals in Biology, Vol. VI, W.A. Pryor, Ed., Academic Press, Inc., Orlando, FL., pp. 199-209).

The foregoing observations lead to the hypothesis that part of the damage observed in irradiated normal cells, tissues, and organs is due to infiltration of host leukocytes that produce toxic free radicals, e.g., oxygen free radicals. In accordance with the present invention, it has now been found that administration of compounds that suppress or detoxify free radicals generated by leukocytes after irradiation would result in reduction of damage to irradiated tissues and organs. Research in the inventors' laboratory has demonstrated that nitroxides are highly efficient protectors against superoxide and hydrogen peroxide cytotoxicity (Mitchell et al. (1990) Biochemistry 29(11):2802-2807). It has now been discovered that if administered during the time period in which leukocytes enter normal tissues and organs after radiation treatment, nitroxides should provide protection against radicals produced by leukocytes, thereby decreasing the damage mediated by such radicals to tissues and organs.

This hypothesis has been tested in experiments where both kidneys of mice were locally irradiated with single, acute doses of radiation. Immediately following irradiation, one group of mice received the nitroxide TEMPOL in their drinking water *ad lib* (final TEMPOL concentration in water = 10 mg/ml) for two weeks post-irradiation. Following the two week TEMPOL treatment, the animals

were returned to regular water, and followed for 28 weeks, with monitoring for weight gain. At 28 weeks after irradiation, the animals were sacrificed, and histological analysis was performed on their kidneys to assess damage. The results are shown in Figures 10 and 11.

Figure 10 shows the mean weight gain over 28 weeks post-irradiation for four groups of animals (20 animals per group). Control = no treatment (closed triangles); animals receiving no radiation, but given 10 mg/ml TEMPOL in their drinking water for two weeks (open triangles); animals receiving 24 Gy to the kidneys (closed circles); animals receiving 24 Gy to the kidneys plus 10 mg/ml TEMPOL in their drinking water two weeks post-irradiation (open triangles).

While mean weight gain is not a direct indicator of kidney damage, it is instructive with respect to overall toxicity of a given treatment. The data in Figure 10 clearly show that animals receiving 24 Gy to the kidneys did not gain weight compared to unirradiated control animals. In fact, weight gain was minimal in the 24 Gy group over the 28 week period.

In contrast, the weight gain in animals given TEMPOL alone was similar to that in control animals. Likewise, animals that received 24 Gy to the kidneys plus TEMPOL gained weight similar to that in the controls.

Based on this endpoint of toxicity, it can be seen that TEMPOL provided radiation protection to the animals.

A more direct assay to determine kidney damage is histological

evaluation. Figure 11 shows histologic sections from mouse kidneys taken 28 weeks after radiation treatment.

Figure 11A shows a section from a kidney of a control animal not treated with radiation. Sections from kidneys of animals receiving TEMPOL alone were identical to those of controls (data not shown).

Figure 11B shows a section of a kidney from a mouse treated with a single radiation dose of 24 Gy. Significant evidence of sclerotic glomeruli and tubular cell loss can be seen.

Figure 11C shows a section of a kidney from a mouse that received a single radiation dose of 24 Gy and orally administered TEMPOL for two weeks post-irradiation. Protection against glomerular sclerosis and tubular cell loss can be readily observed.

These results clearly demonstrate that the representative nitroxide TEMPOL provided significant protection against radiation-induced organ damage after radiation exposure. This may be due to modulation of the cytotoxic activity of infiltrated leukocytes by TEMPOL.

In one aspect of the present invention, radiation-induced damage to normal tissues and organs, including kidney, heart, gut, lung, liver, brain, etc., in which radiation induces inflammation, can be prevented or treated by administering any of the metal-independent nitroxides or oxazolidine compounds that can form an oxazolidine-1-oxyl disclosed above after radiation treatment. These metal-independent nitroxides or oxazolidine compounds can be administered by intermittent or continuous i.v. infusion, or by

intramuscular, subcutaneous, or oral administration. The amount of nitroxide or oxazolidine compound administered can be in the range of from about 1mg/kg body weight/day to about 10 gm/kg body weight/day, more preferably in the range of from about 10 mg/kg body weight/day to about 5 gm/kg body weight/day, and most preferably in the range of from about 25 mg/kg body weight/day to about 2.5 gm/kg body weight/day. A typical dose could be in the range of from about 50 mg/kg body weight/day to about 150 mg/kg body weight/day, preferably 100 mg/kg body weight/day. The duration of treatment can be in the range of from about 1 week to about 24 weeks, and treatment can be begun within about one-half hour to about 24 hours after radiation exposure.

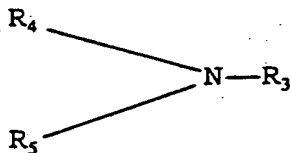
Pharmaceutical compositions useful for treating or preventing damage after radiation exposure can comprise an anti-radiation damage effective amount of a metal-independent nitroxide, an oxazolidine compound capable of forming an oxazolidine-1-oxyl, or a physiologically acceptable salt of either of the foregoing, and a biologically acceptable carrier, wherein the effective amount of the compound is sufficient to protect normal cells, tissues, and organs from damage after exposure thereof to radiation.

The invention being thus described, it will be obvious that the same may be varied in many ways. Such variations are not to be regarded as a departure from the spirit and scope of the invention, and all such modifications as would be obvious to one skilled in the art are intended to be included within the scope of the following claims.

What is claimed is:

1. A pharmaceutical composition comprising an anti-oxidative stress effective amount of a compound selected from the group consisting of a metal-independent nitroxide, an oxazolidine compound capable of forming an oxazolidine-1-oxyl, and a physiologically acceptable salt of either of the foregoing, and a biologically acceptable carrier, wherein said effective amount of said compound is sufficient to protect biological material from oxidative stress.

2. The pharmaceutical composition of claim 1, wherein the compound has the formula:



wherein R_3 is selected from the group consisting of $-O\cdot$ and $-OH$, and R_4 and R_5 combine together with the nitrogen to form a heterocyclic group, or

wherein R_4 and R_5 themselves comprise a substituted or unsubstituted cyclic or heterocyclic group,

or a physiologically acceptable salt thereof.

3. The pharmaceutical composition of claim 2, wherein said heterocyclic group is selected from the group consisting of a

piperidine, pyrrole, imidazole, oxazole, thiazole, pyrazole, 3-pyrroline, pyrrolidine, pyridine, pyrimidine, purine, and a derivative thereof.

4. The pharmaceutical composition of claim 1, wherein said compound is present in an amount of from 25% - 50% by weight of the total composition.

5. The pharmaceutical composition of claim 1, wherein said compound is present in an amount of from 1% to 90% by weight of the total composition.

6. The pharmaceutical composition of claim 1, wherein the compound is 4-hydroxy-2,2,6,6-tetramethylpiperidine-1-oxyl and the compound is present in an amount of from about 0.01 to about 100 mg/ml of the total composition.

7. A method for treating the effects of oxidative stress due to the production of harmful free radical species, comprising administering a composition comprising an anti-oxidative stress effective amount of a compound selected from the group consisting of a metal-independent nitroxide, an oxazolidine compound capable of forming an oxazolidine-1-oxyl, and a physiologically acceptable salt thereof to an organism or biological material susceptible to oxidative stress.

8. The method of claim 7, wherein the oxidative stress is due to the formation of free radicals by an oxidizing agent, oxygen-induced degeneration or disease, ionizing radiation, a carcinogenic agent, a chemotherapeutic agent, a mutagenic agent, aging, or arthritis.

9. The method of claim 7, wherein the oxidative stress is due to reperfusion injury.

10. The method of claim 7, wherein the oxidative stress is due to increased oxygen exposure associated with pulmonary adult respiratory distress syndrome.

11. The method of claim 7, wherein the effect of oxidative stress is oxygen-induced lenticular degeneration, cataracts, or hyaline membrane disease in infants.

12. The method of claim 7, wherein the compound is administered parenterally, intramuscularly, subcutaneously, intravenously, intra-articularly, transdermally, orally, buccally, as a suppository or pessary, or as an aerosol spray or drops.

13. The method of claim 7, wherein the compound is 4-hydroxy-2,2,6,6-tetramethylpiperidine-1-oxyl, and the anti-oxidative stress effective amount is from about 0.1 to about 300 mg/kg/day orally or by intravenous injection, or about 0.1 to about

200 mg/kg/day by inhalation.

14. A method of prolonging the storage life of human or animal cells, tissues, or organs *in vitro*, comprising contacting said cells, tissues, or organs *in vitro* with a solution containing an anti-oxidative stress effective amount of a compound selected from the group consisting of a metal-independent nitroxide, an oxazolidine compound capable of forming an oxazolidine-1-oxyl, and a physiologically acceptable salt thereof to protect said cells, tissues, or organs from oxidative stress.

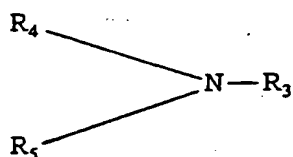
15. A pharmaceutical composition comprising an anti-radiation damage effective amount of a compound selected from the group consisting of a metal-independent nitroxide, an oxazolidine compound capable of forming an oxazolidine-1-oxyl, and a physiologically acceptable salt of either of the foregoing, and a biologically acceptable carrier, wherein said effective amount of said compound is sufficient to protect normal cells, tissues, and organs from damage after exposure thereof to radiation.

16. The pharmaceutical composition of claim 15, wherein said compound is 4-hydroxy-2,2,6,6,-tetramethylpiperidine-1-oxyl.

17. A method for treating or preventing damage to normal cells, tissues, or organs in a mammal that has been exposed to ionizing radiation, comprising administering to said mammal, after

exposure to ionizing radiation, a composition comprising an anti-radiation damage effective amount of a compound selected from the group consisting of a metal-independent nitroxide, an oxazolidine compound capable of forming an oxazolidine-1-oxyl, and a physiologically acceptable salt thereof.

18. The method of claim 17, wherein said compound has the formula:



wherein R_3 is selected from the group consisting of $-O\cdot$ and $-OH$, and R_4 and R_5 combine together with the nitrogen to form a heterocyclic group, or

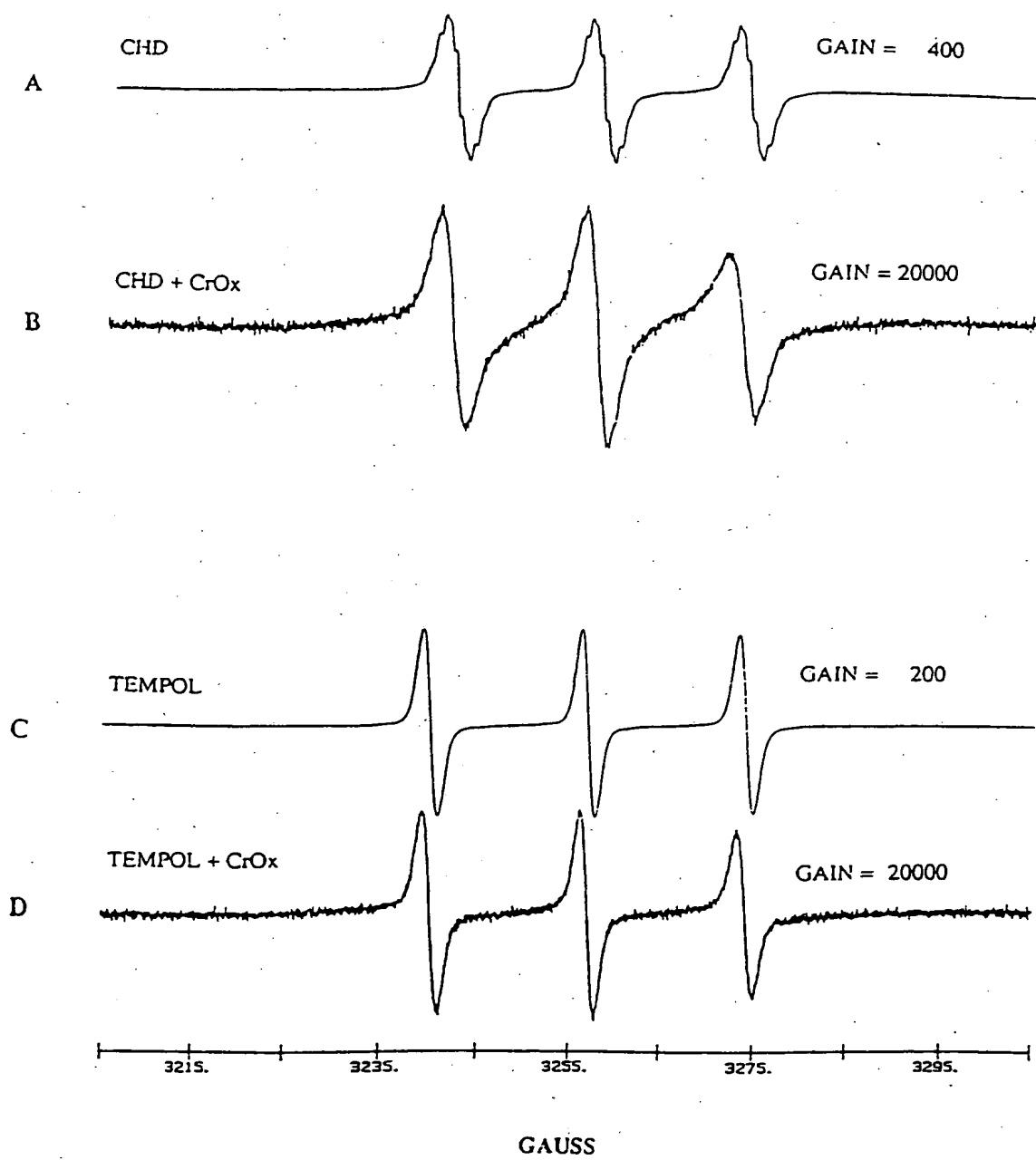
wherein R_4 and R_5 themselves comprise a substituted or unsubstituted cyclic or heterocyclic group,

or a physiologically acceptable salt thereof.

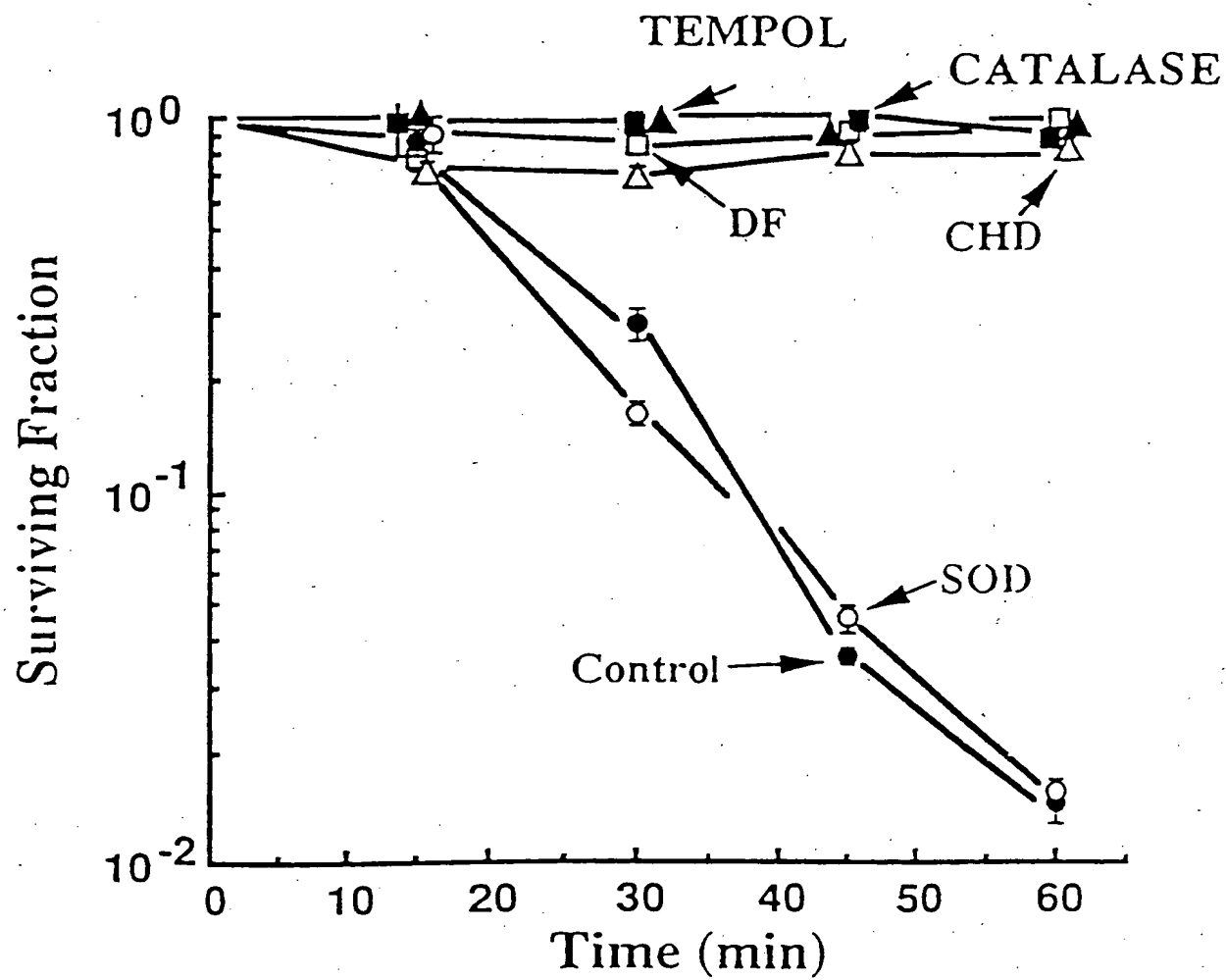
19. The method of claim 17, wherein said compound is 4-hydroxy-2,2,6,6-tetramethylpiperidine-1-oxyl.

20. The method of claim 17, wherein said administering is begun within a period of from about one-half hour to about 24 hours after exposure of said mammal to radiation.

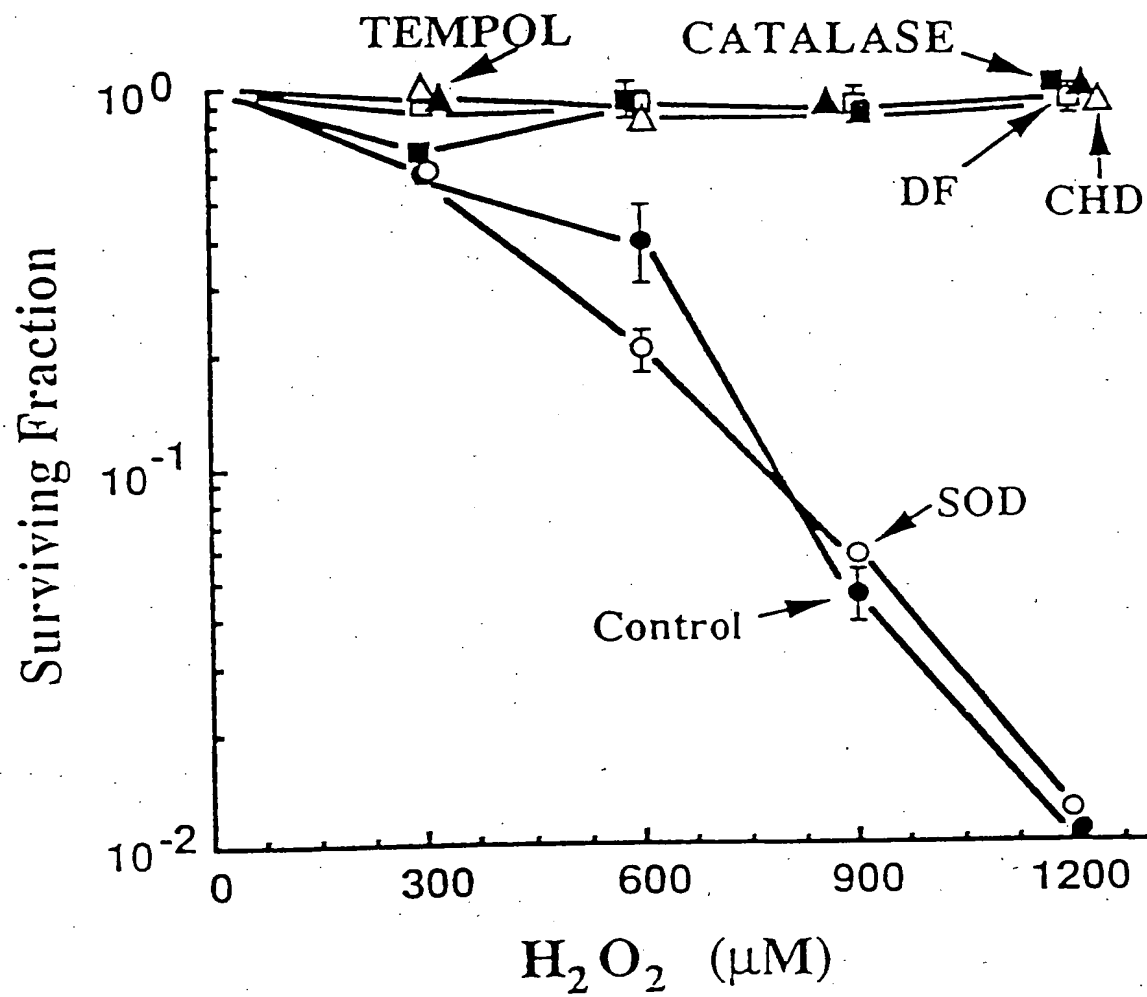
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FIGURE 1

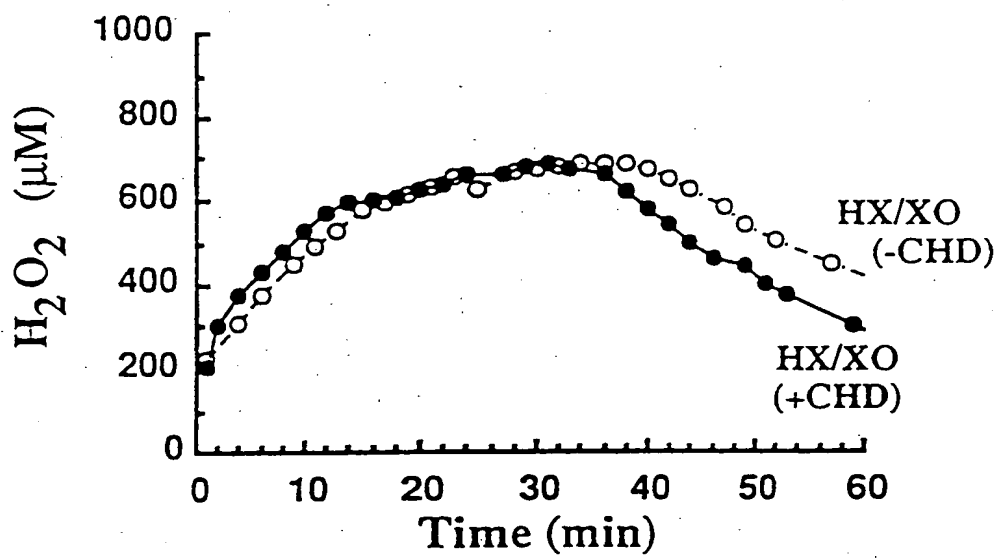
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FIGURE 2

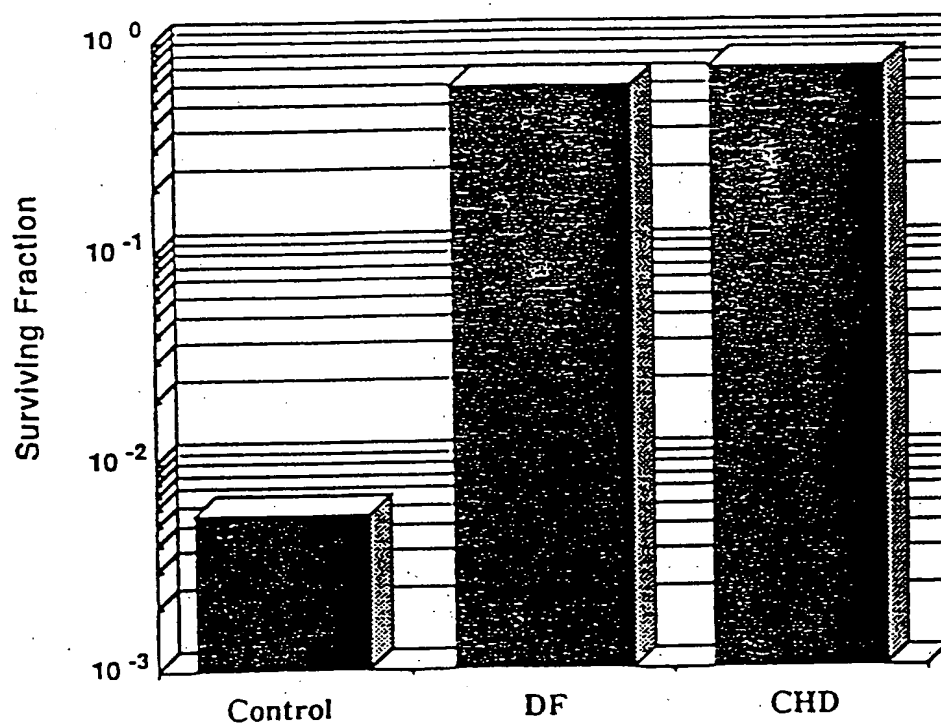
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FIGURE 3

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FIGURE 4

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FIGURE 5

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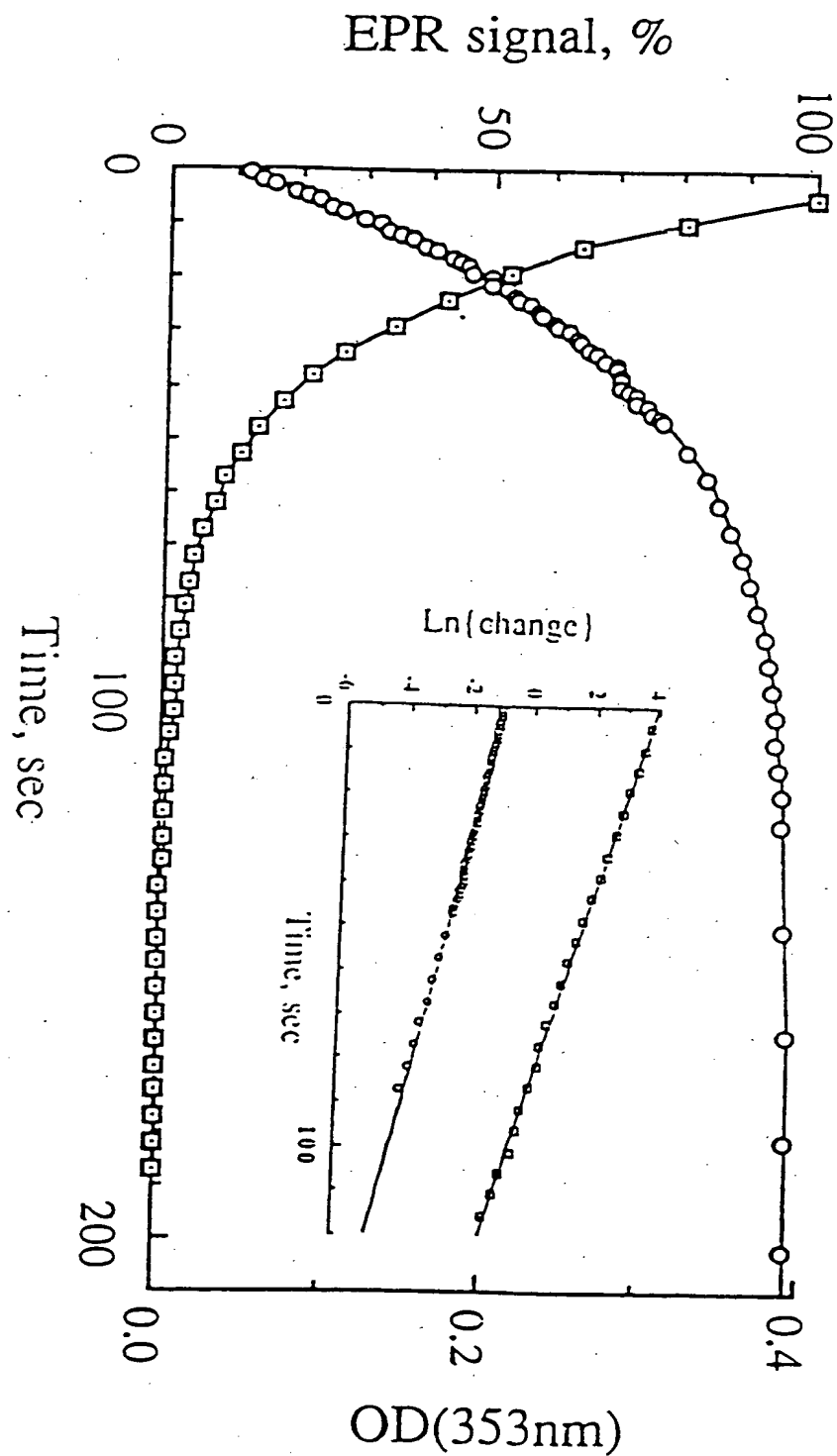


FIGURE 6

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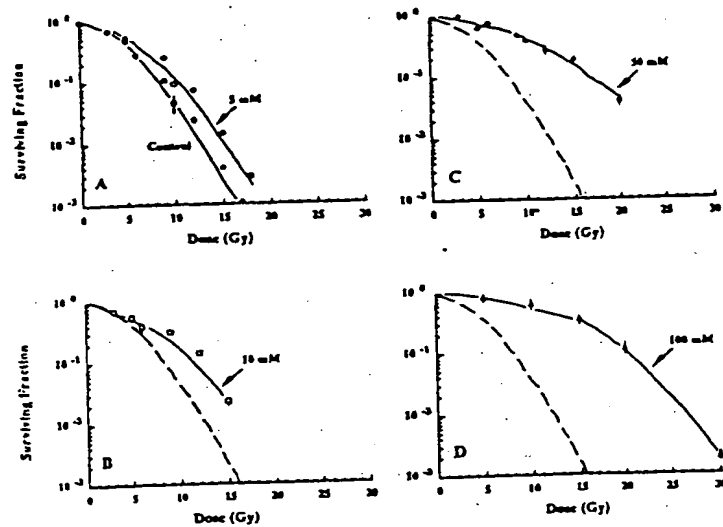


FIGURE 7

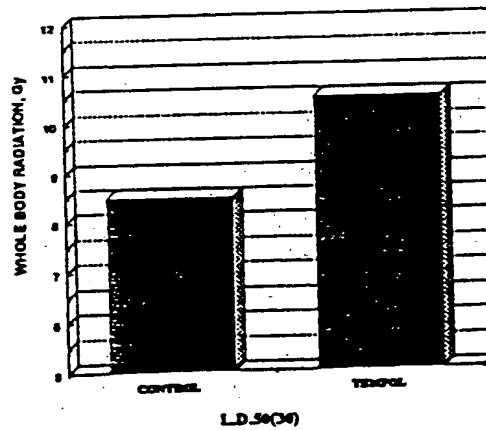
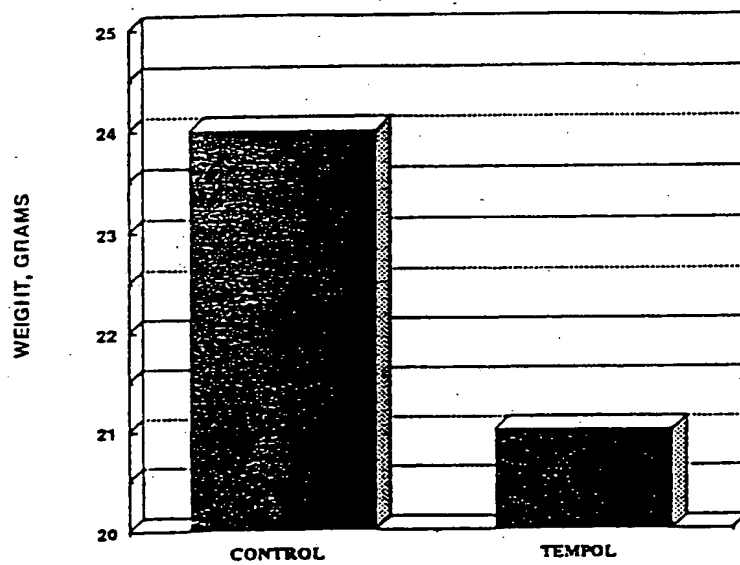


FIGURE 8

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FIGURE 9

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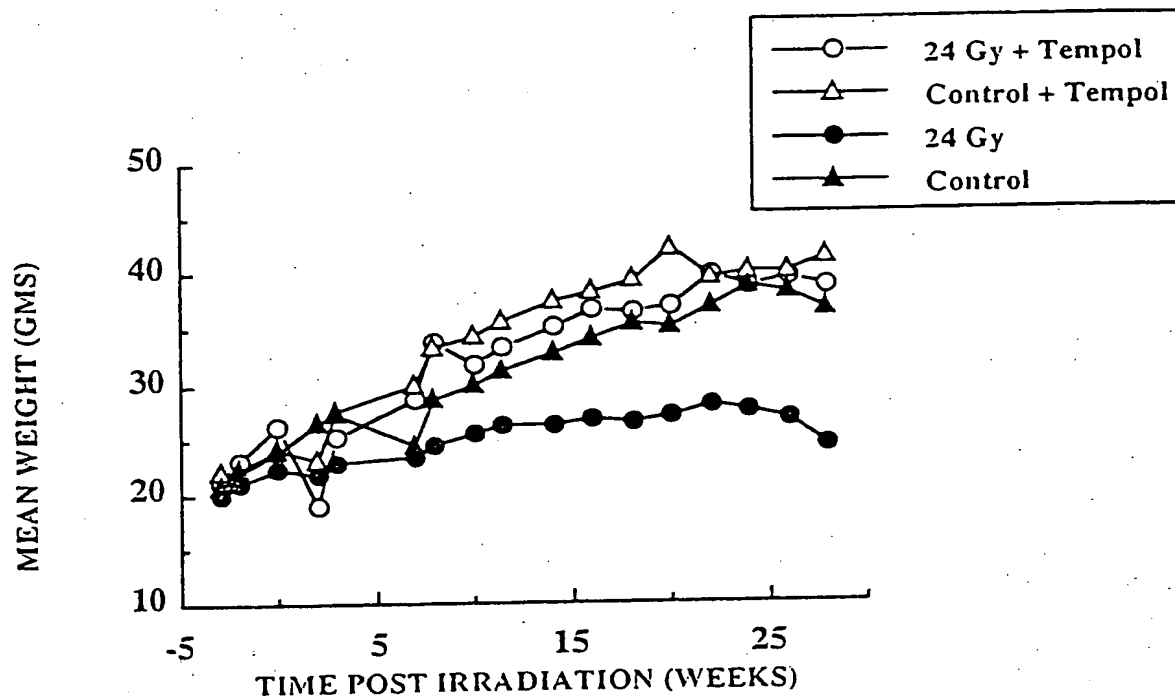
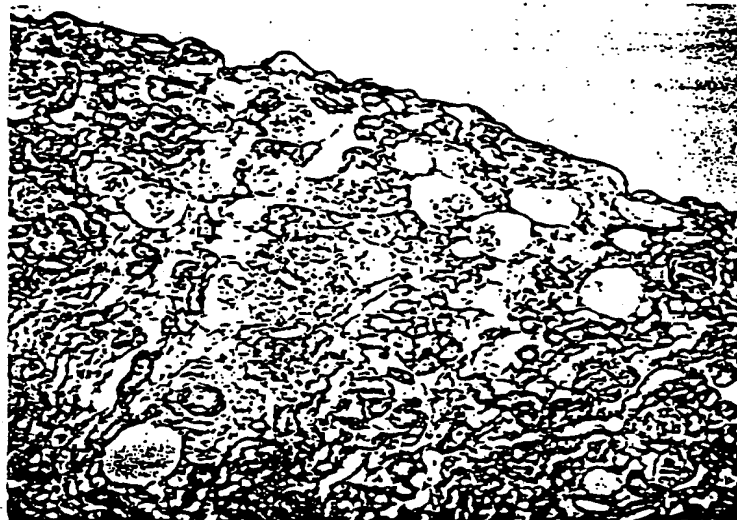


FIGURE 10

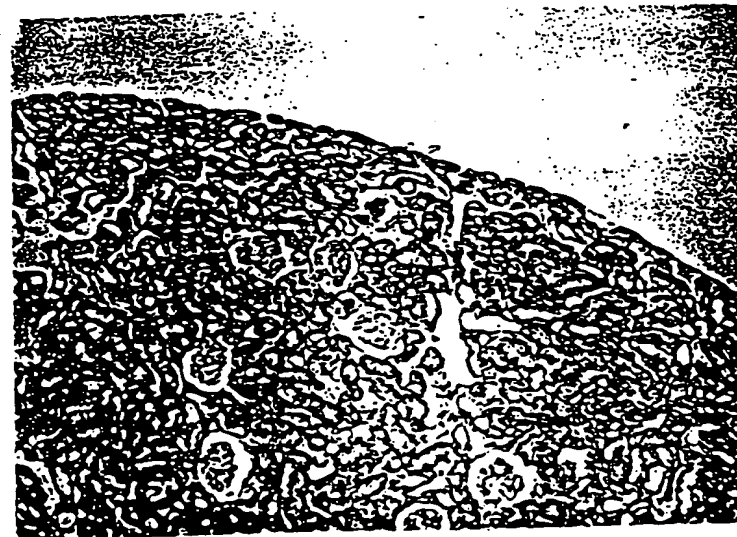
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A



B



C

FIGURE 11

INTERNATIONAL SEARCH REPORT

Inter national Application No
PCT/US 96/09524

A. CLASSIFICATION OF SUBJECT MATTER
IPC 6 A61K31/42 A61K31/445

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
IPC 6 A61K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO 91 13619 A (US GOVERNMENT) 19 September 1991 see claims 1-20; example 4; table 1B ---	1-20
X	WO 88 05044 A (PHARMACIA AB) 14 July 1988 see example 1; table 1 ---	1-13
X	WO 88 05653 A (PROCTOR PETER H) 11 August 1988 see page 8, line 30 - page 9, line 15 ---	1-13
X	WO 95 05397 A (HSIA JEN CHANG) 23 February 1995 see page 23 see page 32, last paragraph - page 36 -----	14

☐ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

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European Patent Office, P.B. 5818 Patentlaan 2
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Foerster, W

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Information on patent family members

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